

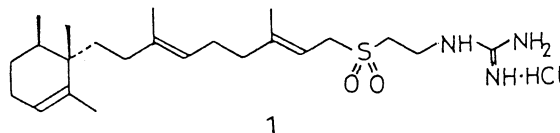
Total Synthesis of (+)-Agelasidine C,  
a Physiologically Active Marine Diterpenoid with Hypotaurocyamine Group<sup>1)</sup>

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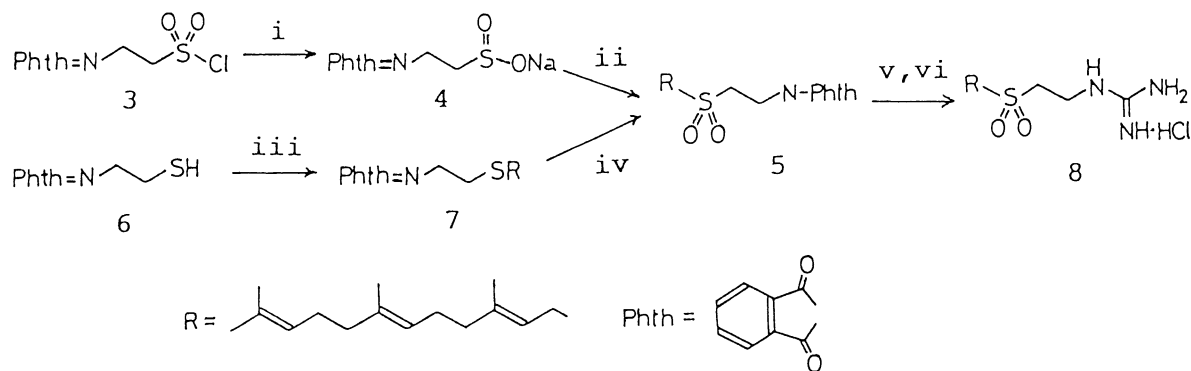
The introduction of hypotaurocyamine unit to terpenic chain was examined using farnesyl bromide as a model compound and this was feasible by chemoselective oxidation of the derived sulfide by peracid. By the application of the method thus established, a total synthesis of agelasidine C has been accomplished.

Agelasidines are a group of physiologically active constituents found in marine sponges of *Agelas* species and structurally characterized by the presence of novel hypotaurocyamine unit as a polar functionality.<sup>2)</sup> In continuation of the synthetic studies on *Agelas* terpenoids,<sup>3)</sup> we investigated the synthesis of agelasidine C (**1**), of which monocyclic diterpenoid moiety was recently synthesized by us.<sup>4)</sup>



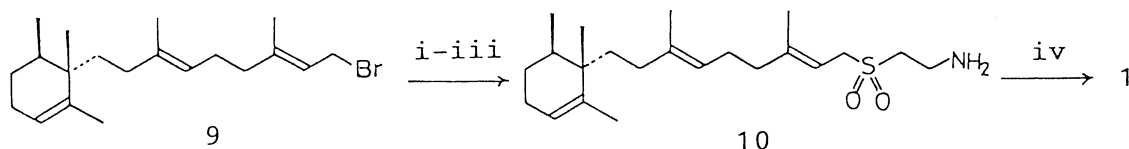
The installation of the hypotaurocyamine group to the terpenic unit was studied using farnesyl bromide (**2**) as a model compound (Scheme 1). The first approach was the substitution reaction of **2** with sodium phthalimidoethanesulfinate (**4**) to give the sulfone directly. The reaction of **2** with the crude sulfinate salt **4**, prepared by the reduction of the corresponding sulfonyl chloride,<sup>5)</sup> afforded **5** in 23% yield. The low yield and the lack of reproducibility in this procedure, mainly attributable to the problems associated in purification of **4**, led us to the examination of second approach. Namely, we investigated a route *via* sulfide **7**, where the chemoselective oxidation to the sulfone **5** was mandatory. The compound **7** was prepared by the alkylation of the thiol **6** with **2**. Of the several methods examined for the oxidation of **7** to **5**, the satisfactory result was obtained by the oxidation with *m*-chloroperbenzoic acid (MCPBA), in which the solvent effect was remarkable. Whereas the reaction with MCPBA (2 equiv) in CH<sub>2</sub>Cl<sub>2</sub> afforded **5** in 16% yield after removal of the epoxidized products, the reactions in etheric solvents gave **5** in higher yields (Et<sub>2</sub>O, 54%; THF, 66%).<sup>6)</sup> The sulfone **5** was deprotected and the guanylation of the product amine by treatment with 2-methyl-2-pseudothiourea sulfate gave the hypotaurocyamine derivative **8** in 85% yield.

Having established the route to introduce the hypotaurocyamine unit on a terpenic chain, we proceeded to the synthesis of agelasidine C starting from the diterpenic bromide **9**<sup>4,7)</sup> (Scheme 2). The peracid oxidation of the derived sulfide



Scheme 1. i) Zn, H<sub>2</sub>O, then NaOH, Na<sub>2</sub>CO<sub>3</sub>; ii) **2**, DMF; iii) **2**, Et<sub>3</sub>N, THF; iv) MCPBA, THF; v) NH<sub>2</sub>NH<sub>2</sub>, EtOH; vi) NH<sub>2</sub>C(=NH)SMe·½H<sub>2</sub>SO<sub>4</sub>, H<sub>2</sub>O

afforded the corresponding sulfone in a yield higher than in the model case, of which deprotection provided the hypotaaurin derivative **10**. The reaction of **10** with 2-methyl-2-pseudothiourea resulted in the formation of the hypotaurocyamine compound only in low yield (10-18%), probably due to the low solubility of the substrate **10** in water. However when the reaction performed with the use of 3,5-dimethylpyrazole-1-carboxamide nitrate,<sup>8)</sup> the aimed product was obtained in an acceptable yield. The spectral data (IR, <sup>1</sup>H and <sup>13</sup>C NMR) of the synthetic product<sup>9)</sup> were identical with those of the natural product and thus the total synthesis of agelasidine C has been accomplished.<sup>10)</sup>



Scheme 2. i) **6**, Et<sub>3</sub>N, THF (51%); ii) MCPBA, THF (82%); iii) NH<sub>2</sub>NH<sub>2</sub>, EtOH (93%); iv) 3,5-dimethylpyrazole-1-carboxamide nitrate, MeOH (65%)

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- IR(CHCl<sub>3</sub>): ν<sub>max</sub> 3400, 2975, 2940, 2845, 1715, 1680, 1660, 1620, 1450, 1405, 1375, 1300, 1190, 1120, 870, 850 cm<sup>-1</sup>; <sup>1</sup>H NMR(CD<sub>3</sub>OD, 400 MHz): δ 0.86(3H, s), 0.87(3H, d, J = 7.3 Hz), 1.60(3H, br s), 1.62(3H, br s), 1.78(3H, br s), 1.2-2.4(13H, m), 3.31(2H, t, J = 5.5 Hz), 3.72(2H, t, J = 5.5 Hz), 3.90(2H, d, J = 7.9 Hz), 5.11(1H, br s), 5.31(1H, t, J = 7.9 Hz), 5.42(1H, brs); <sup>13</sup>C NMR(CD<sub>3</sub>OD, 100 MHz): δ 16.2, 16.4, 17.1, 19.5, 21.5, 26.5, 27.3, 28.2, 34.6, 35.4, 36.0, 36.5, 40.8, 41.6, 51.3, 54.6, 110.7, 124.0, 125.2, 137.8, 140.7, 148.3, 158.7.
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